



# Development of a mucoadhesive nanoparticulate drug delivery system for a targeted drug release in the bladder

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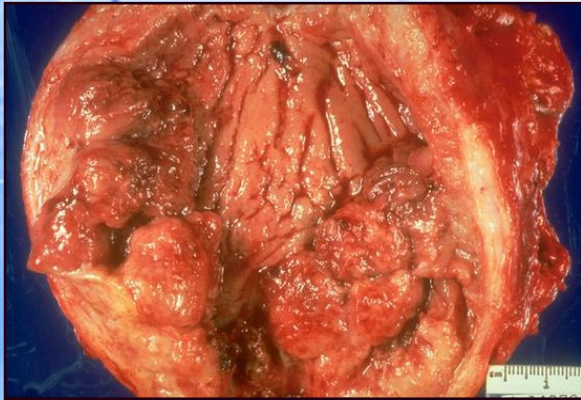
## **1. Introduction**

## **2. Methods & Results**

## **3. Conclusion**

# Introduction

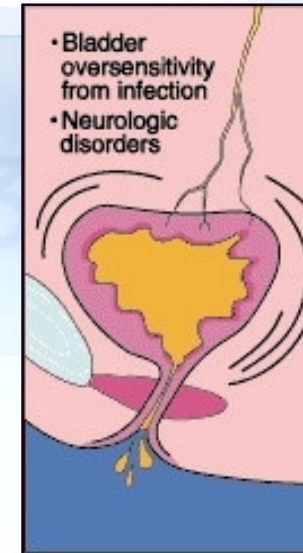
## *Diseases of the urinary bladder*



cancer



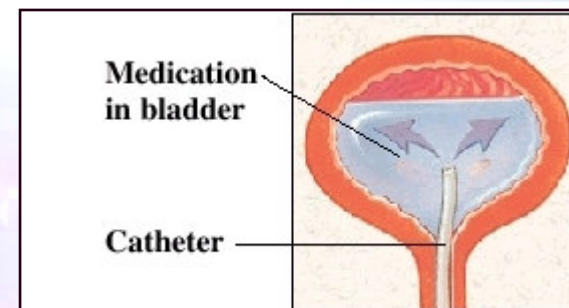
inflammation, infection



incontinence

- treated by oral administration of pharmaceutical compounds  
→ systemic delivery

- Intravesical Drug Delivery (IDD)

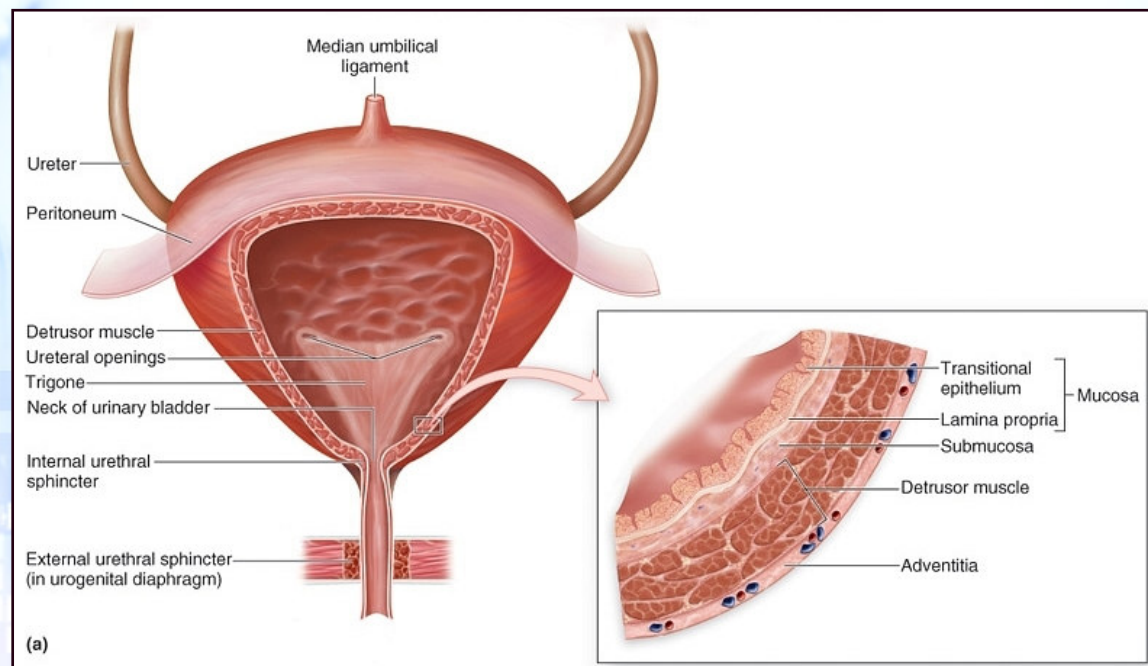




# Introduction

## *Intravesical drug delivery (IDD) limitations*

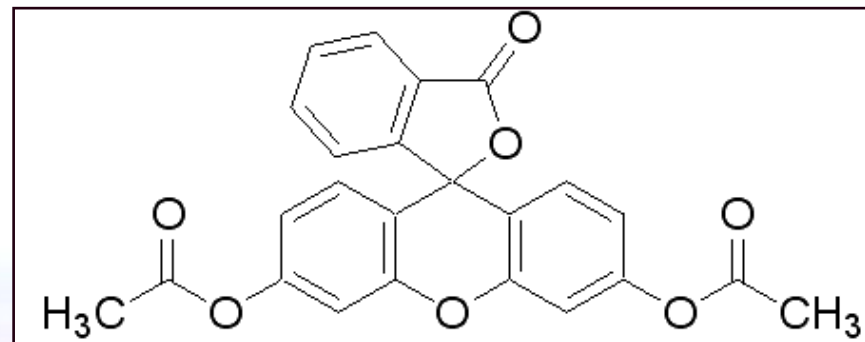
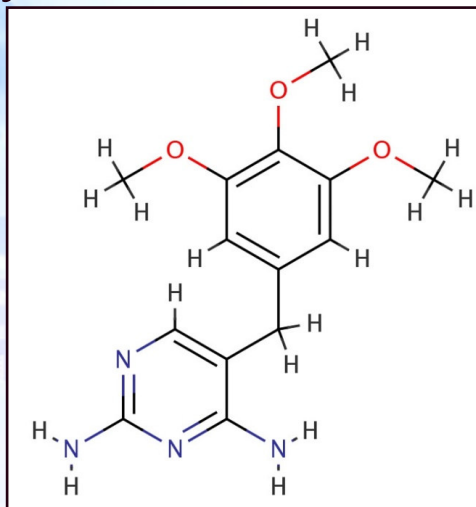
- periodical voiding of urine dilutes and washes out the drug
  - reduces the residence time of drug and lead to a new administration
  - repeated catheterizations increase potential for infections
- very low permeability of the urothelium in the diseased state



# Introduction

## *Purpose of the present study*

- development of a mucoadhesive nanoparticulate drug delivery system for local use in intravesical therapy
  - retarding release of the drug
  - prolong the residence time of the drug in the bladder
- trimethoprim (TMP) was used as an effective local therapy from cystitis in the bladder
- fluorescein diacetate (FDA) was used as fluorescent marker



# Content



1. Introduction

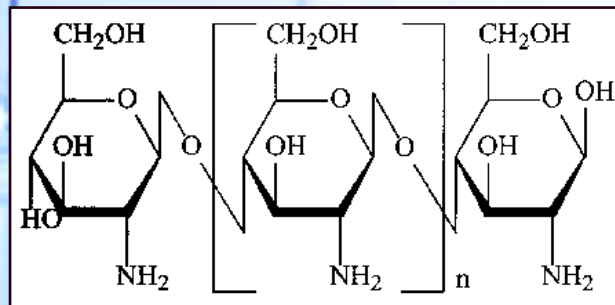
**2. Methods & Results**

3. Conclusion

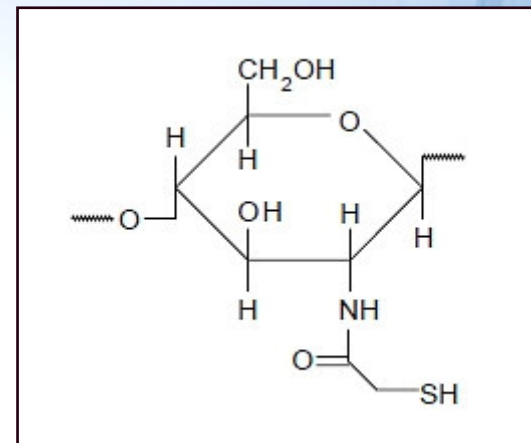


# Methods & Results

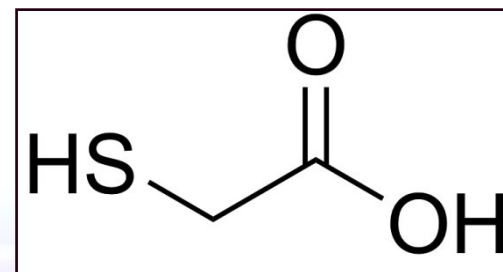
## *Preparation of the matrix of the drug delivery system*



Chitosan



Chitosan - Thioglycolic acid (TGA)



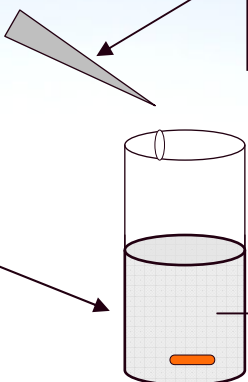
Thioglycolic acid

# Methods & Results

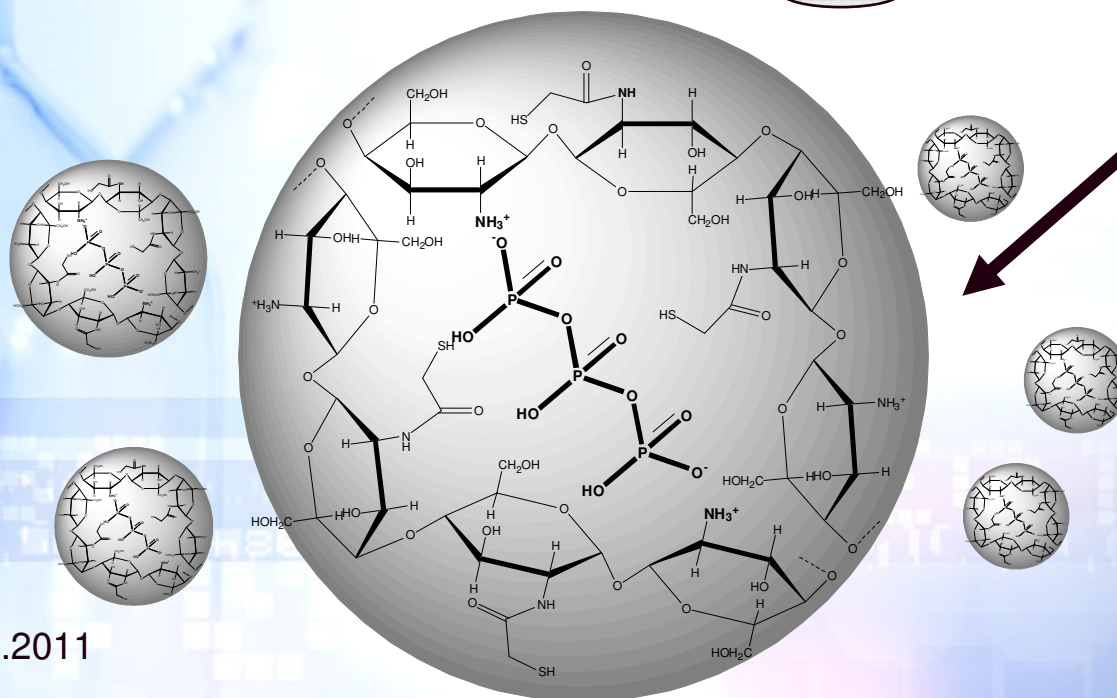
## Preparation of the drug delivery system

Chitosan or chitosan-TGA in  
0.01 M acetic acid /sodium  
acetat buffer 0.5% (w/v) pH 6.2

TPP solution in  
demineralised water  
0.2% (w/v)



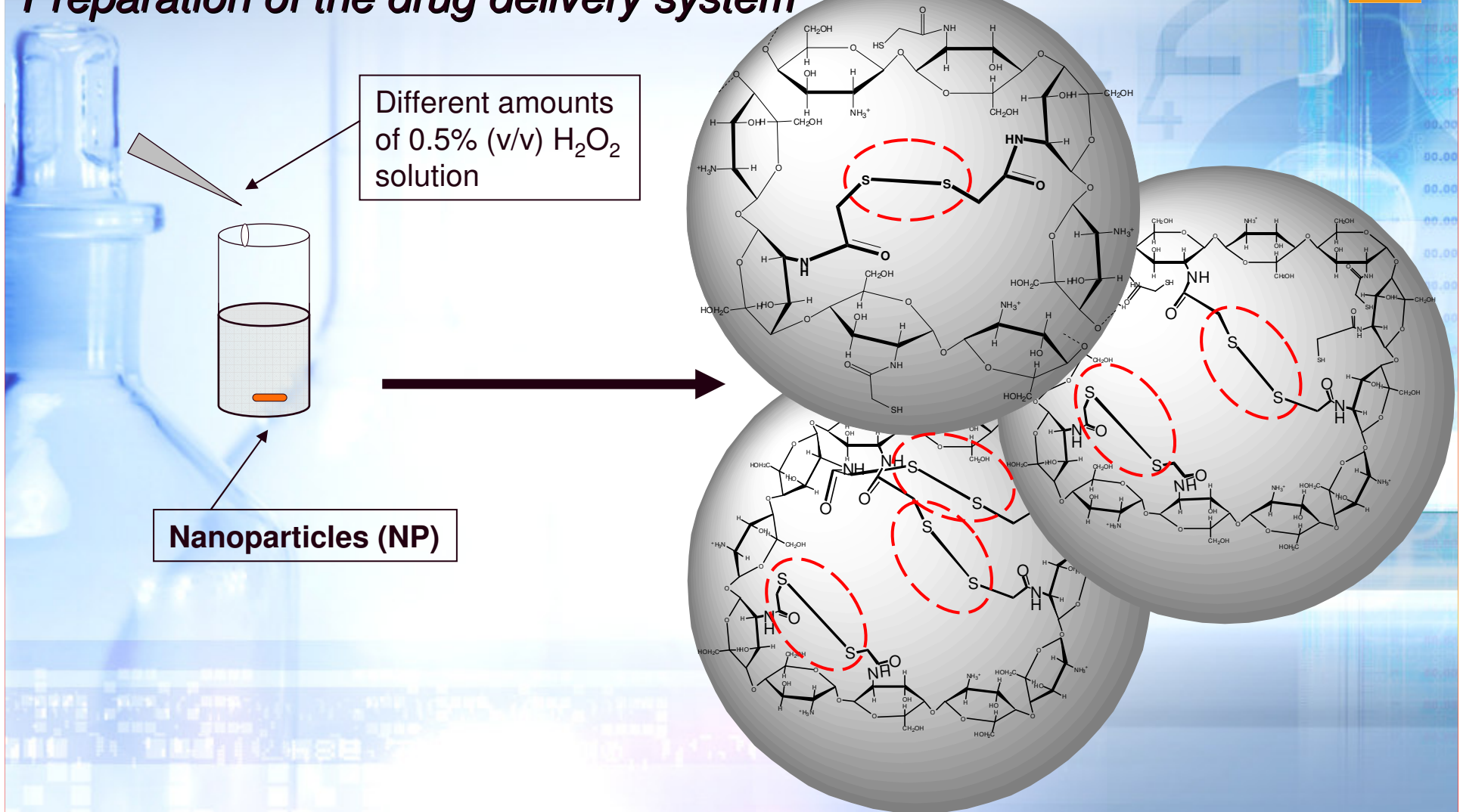
**Nanoparticles (NP)**





# Methods & Results

## Preparation of the drug delivery system



# Methods & Results

## *Characterization of the drug delivery system*

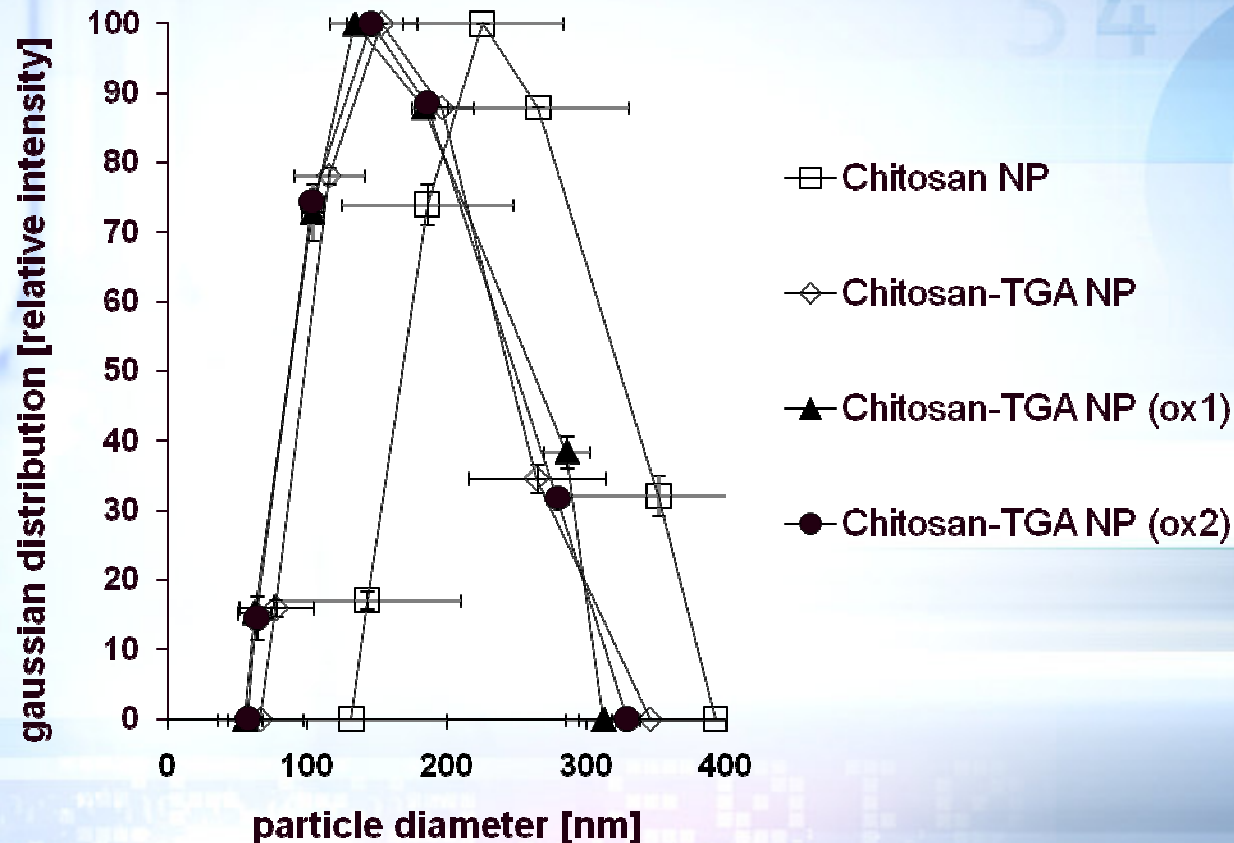
**Table 1.**

Mean particle diameter and zeta potential of chitosan-TGA nanoparticles obtained by ionic gelation with TPP and followed by different oxidation with H<sub>2</sub>O<sub>2</sub>, respectively. Indicated values are means  $\pm$  SD (n  $\geq$  3).

<b>Nanoparticles</b>	<b>Mean particle diameter [nm]</b>	<b>Polydispersity index</b>	<b>Zeta potential [mV]</b>
<b><i>Ionically crosslinked</i></b>			
Chitosan	266 $\pm$ 64	0.44	7 $\pm$ 1
Chitosan-TGA	197 $\pm$ 24	0.38	7 $\pm$ 1
<b><i>Covalently crosslinked</i></b>			
Chitosan TGA (ox1)	183 $\pm$ 7	0.31	13 $\pm$ 3
Chitosan TGA (ox2)	186 $\pm$ 6	0.29	12 $\pm$ 1

# Methods & Results

## Characterization of the drug delivery system



**Fig. 2.** Size distribution of ionically crosslinked nanoparticles as well as covalently crosslinked nanoparticles. Indicated values are means  $\pm$  SD of last three experiments.



# Methods & Results

## *Characterization of the drug delivery system*

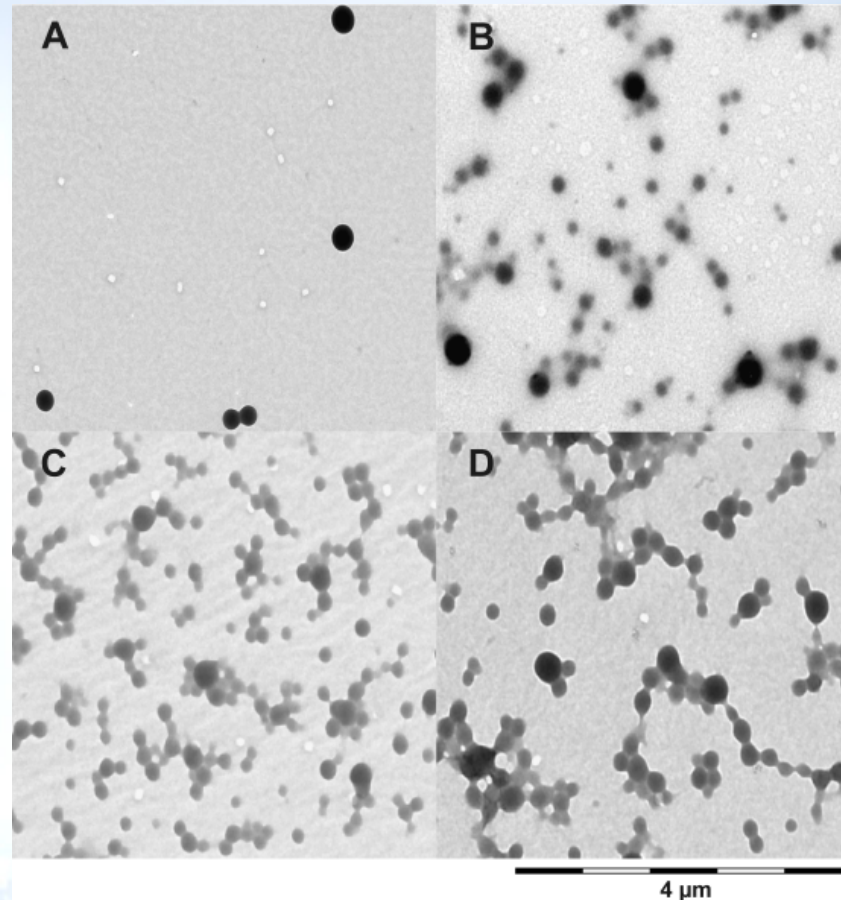
**Table 2.**

Amount of thiol groups and disulfide bonds immobilised on the basic thiomers Chitosan-TGA and nanoparticles after ionic gelation with TPP and different degrees of oxidation with H<sub>2</sub>O<sub>2</sub>, respectively. Indicated values are means ± SD (n ≥ 3).

	H <sub>2</sub> O <sub>2</sub> [μmol]	-SH [μmol/g]	-S-S- [μmol/g]	Σ-SH [μmol/g]
<b>Chitosan-TGA</b>	-	1456	136	1728 ± 62
<b>Chitosan-TGA NP</b>	-	1391	178	1747 ± 36
<b>Chitosan-TGA NP (ox1)</b>	10.60	903	426	1753 ± 55
<b>Chitosan-TGA NP (ox2)</b>	21.21	641	559	1758 ± 27

# Methods & Results

## *Characterization of the drug delivery system*

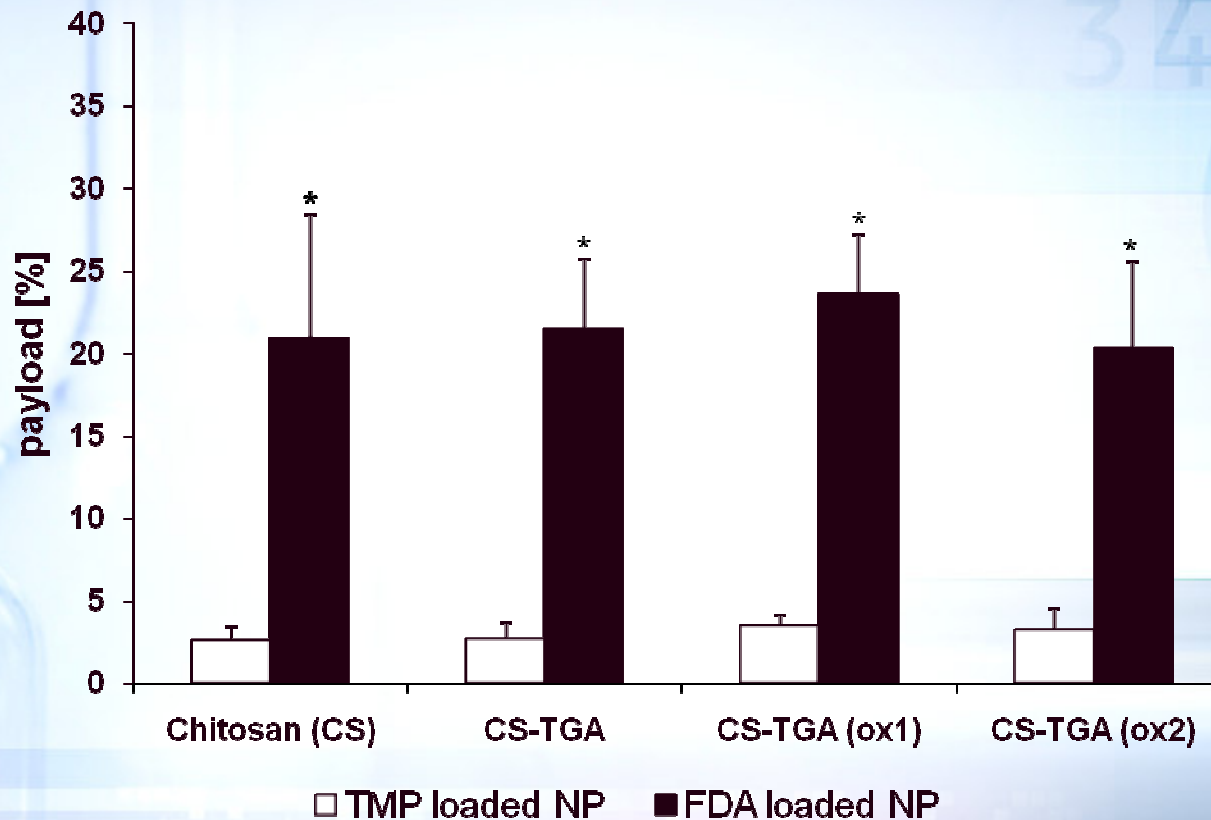


**Fig. 3.** Transmission electron microscopy images of the spherical shape of nanoparticles based on chitosan [A], chitosan-thioglycolic acid [B], chitosan-thioglycolic acid with 426 μmol/g disulfide bonds [C] and chitosan- thioglycolic acid with 559 μmol/g disulfide bonds [D].

20.07.2011 Displayed bar represents 4.0 μm.

# Methods & Results

## Characterization of the drug delivery system



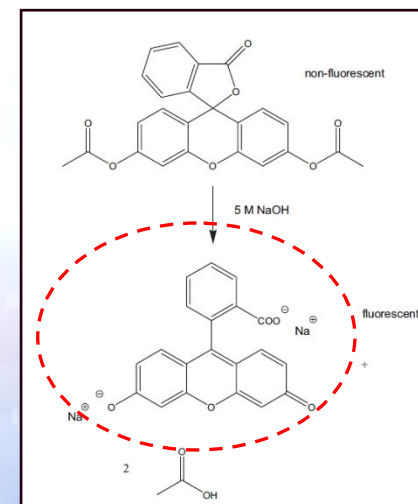
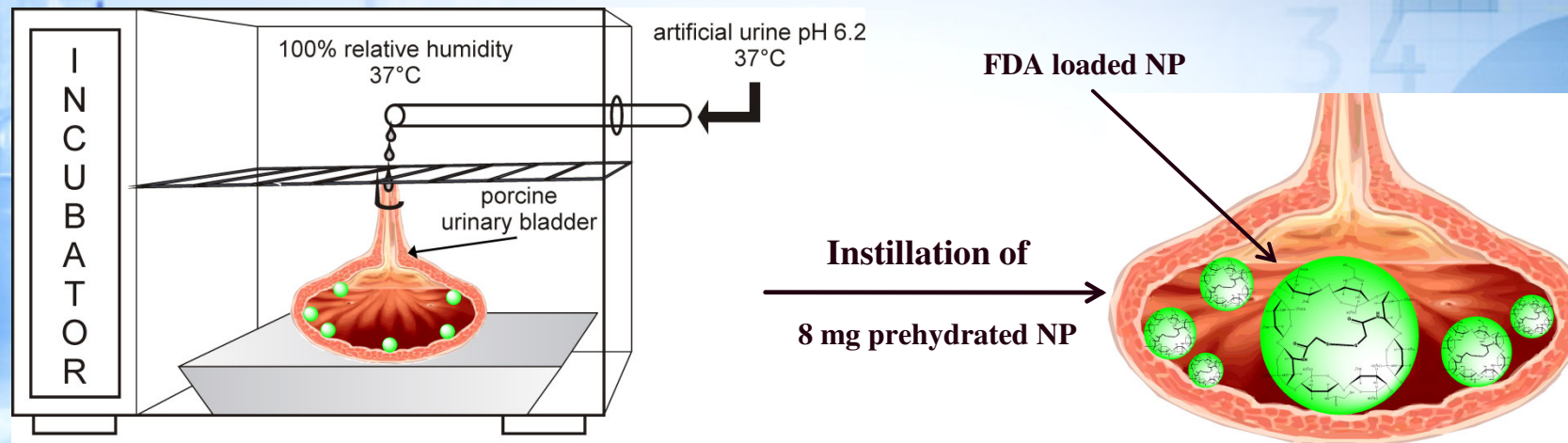
**Fig. 4.** Payload of trimethoprim [white bars] and fluorescein diacetate [black bars] loaded nanoparticles based on chitosan, chitosan-thioglycolic acid and thioglycolic acid with 426  $\mu\text{mol/g}$  (ox1) and 559  $\mu\text{mol/g}$  (ox2) disulfide. Indicated values are means  $\pm$  SD ( $n \geq 3$ ). \* Differs from TMP,  $p < 0.05$ .

chitosan-  
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0.05.



# Methods & Results

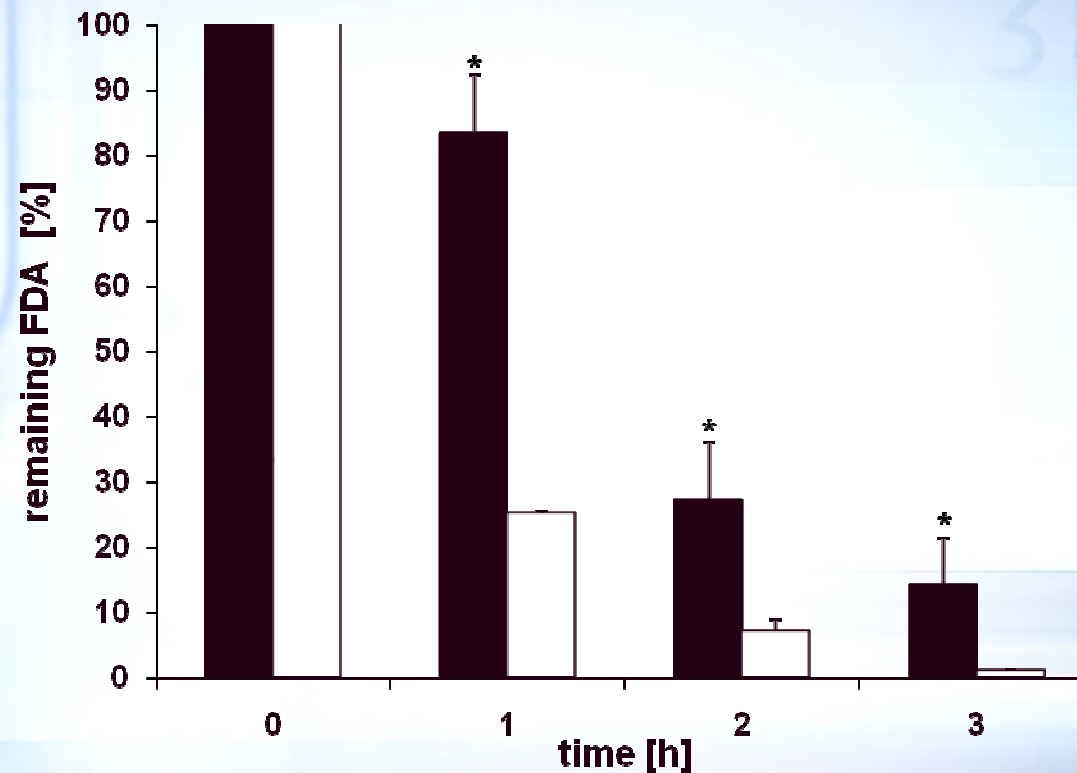
## *In vitro* mucoadhesion studies on porcine urinary bladders



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# Methods & Results

## *In vitro mucoadhesion studies on porcine urinary bladders*



**Fig. 5.** Percentage of fluorescein diacetate remaining on porcine urinary bladders as a function of time. Studies were carried out with chitosan-thioglycolic acid nanoparticles [black bars] and unmodified chitosan nanoparticles [white bars] as control. Indicated values are means  $\pm$  SD ( $n \geq 3$ ). \* Differs from unmodified chitosan nanoparticles,  $p < 0.05$ .

# Methods & Results

## *In vivo evaluation of particles with rats*

- female Sprague-Dawley rats, average body weight 250 g
- rats were fasted but had free access to water
- anesthetized by an injection of ketamine (20 mg/kg)/xylazine-hydrochloride (4 mg/kg) mixture
- before urethral catheterization animals were positioned in supine position, and micturition was induced through mild caudal abdominal massage
- 500  $\mu$ l of each formulation was administered



<b>Animal grouping</b>	<b>Administered formulations</b>
Group 1	FDA suspension
Group 2	FDA loaded unmodified chitosan nanoparticles
Group 3	FDA loaded chitosan-TGA nanoparticles

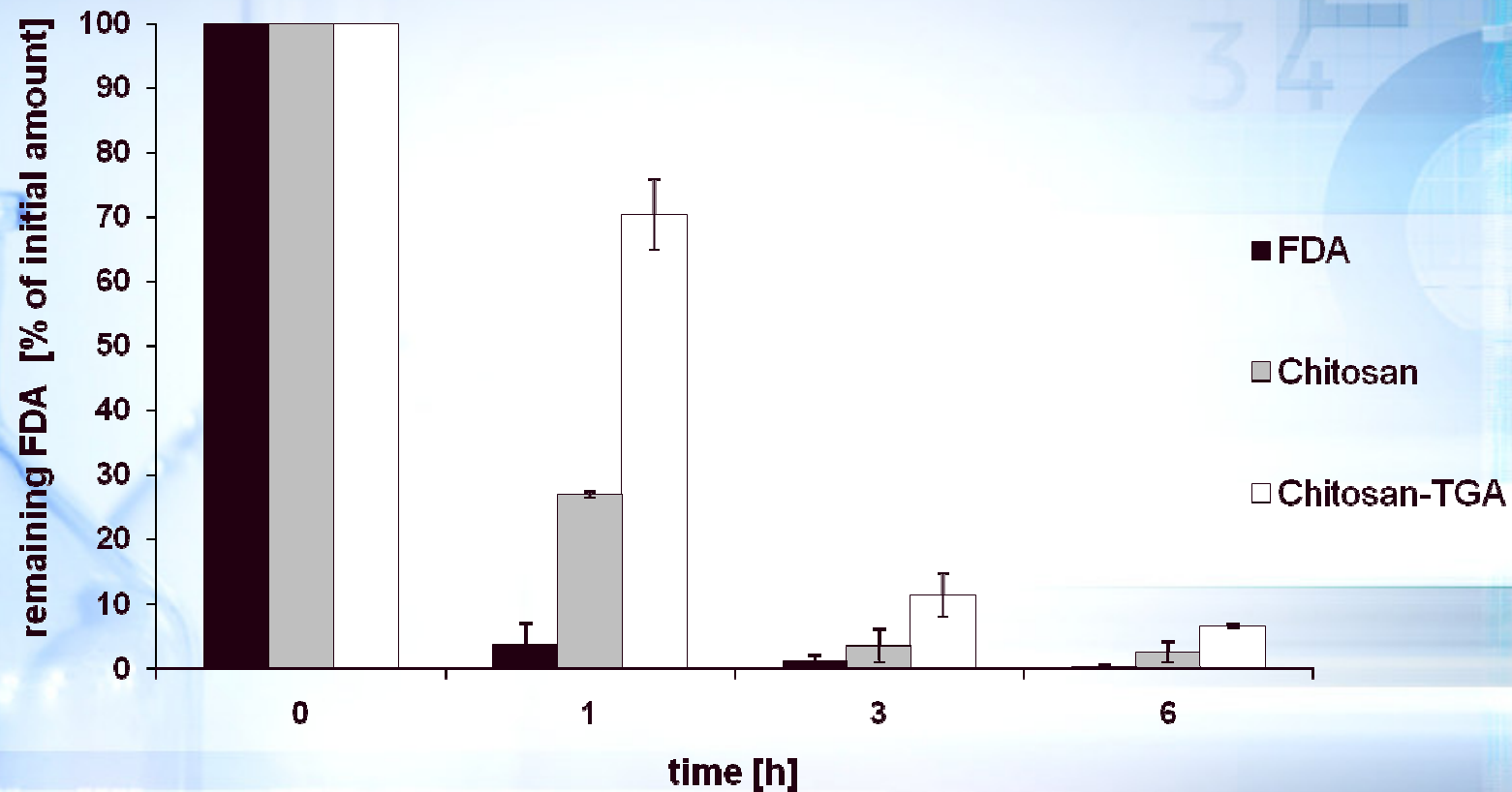


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# Methods & Results

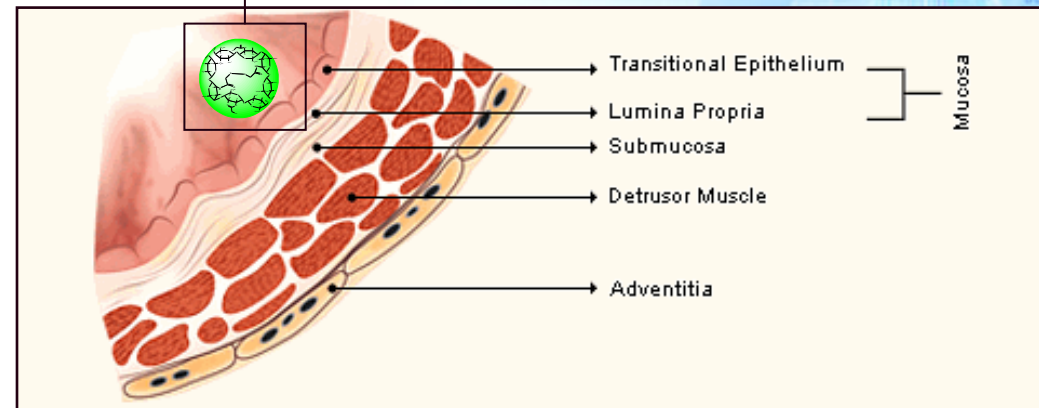
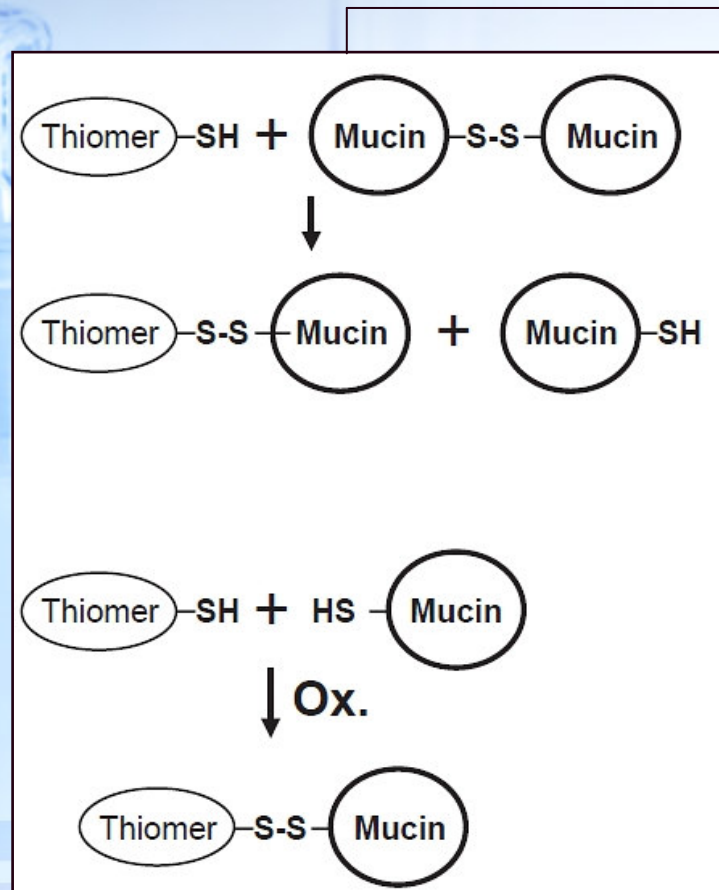
## *In vivo* evaluation of particles with rats



**Fig. 6.** Amount of fluorescein diacetate remaining on rats bladders. Fluorescein diacetate was applied without any excipients [black bars] or incorporated in unmodified chitosan nanoparticles [grey bars] or chitosan-thioglycolic acid nanoparticles [white bars]. Indicated values are means  $\pm$  SD ( $n \geq 3$ ).

# Methods & Results

## *Mucoadhesion on mucosa of urinary bladders*

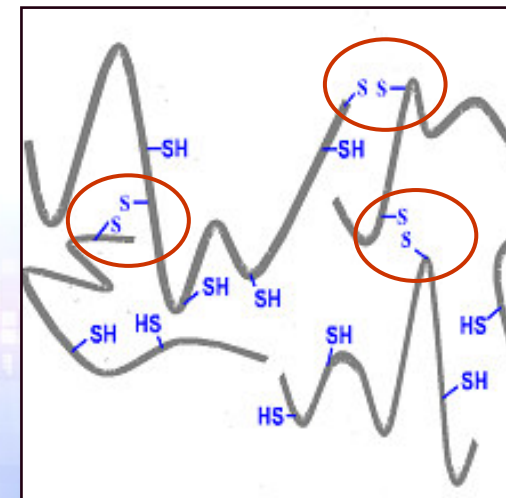
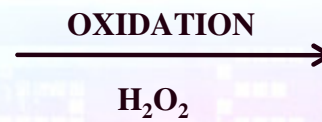
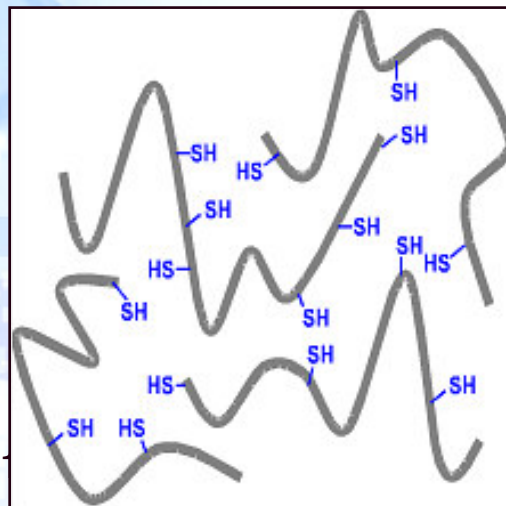
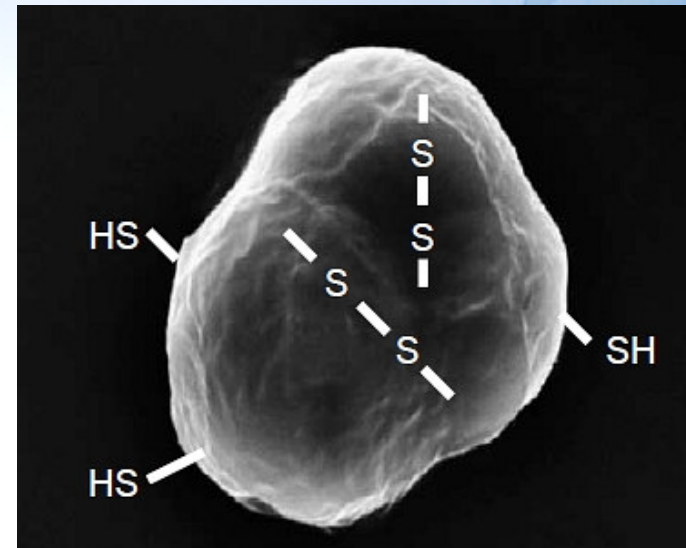
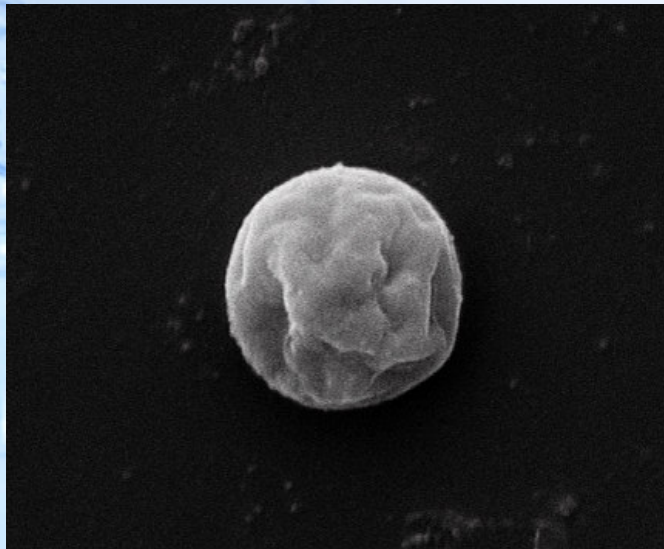


Layers in the bladder wall

Mechanism of disulfide bond formation  
between thiomers and mucus glycoproteins

# Methods & Results

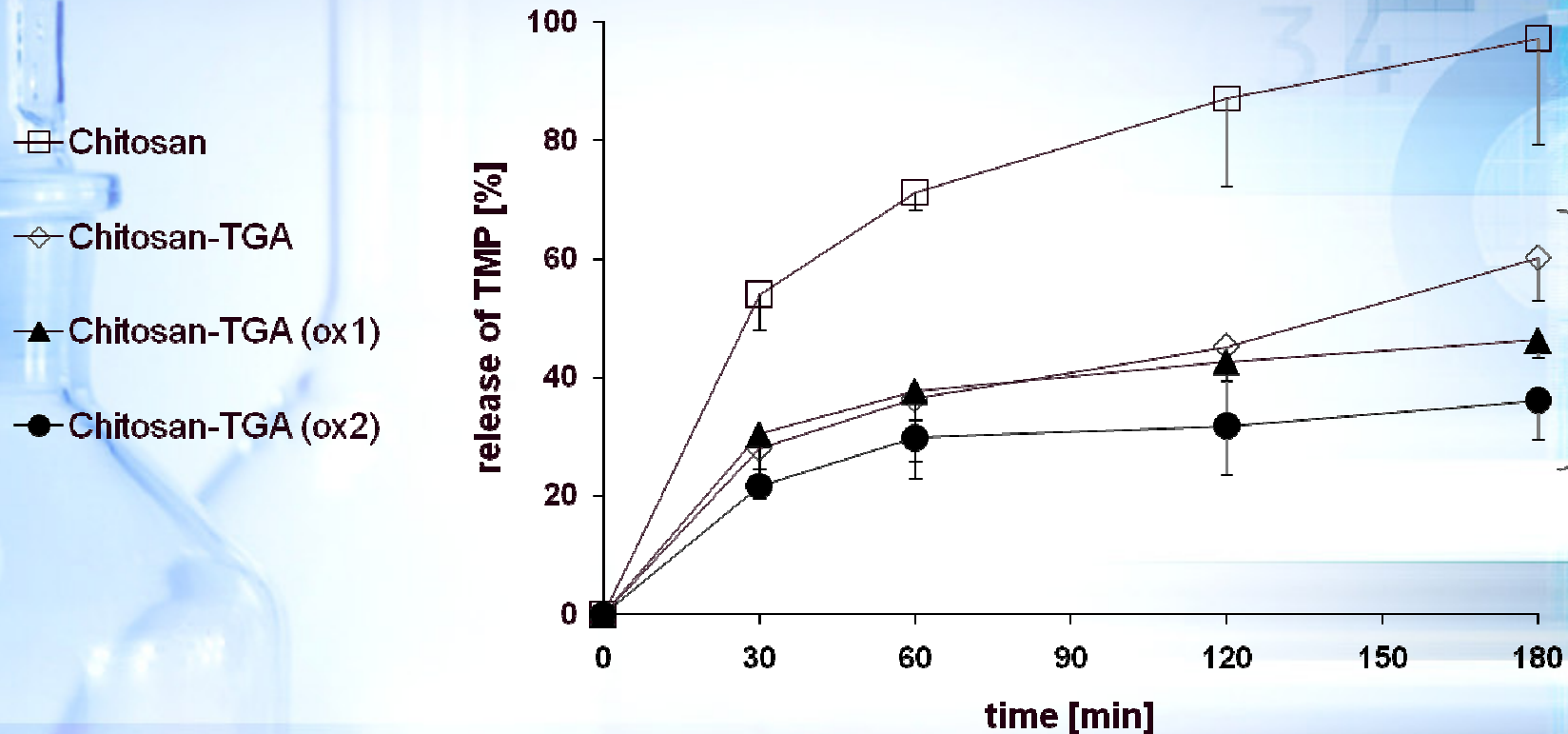
## Release studies





# Methods & Results

## TMP release studies



**Fig. 7.** Release properties of trimethoprim nanoparticles among simulated conditions with artificial urine as a function of crosslinking. Studies were carried out with nanoparticles based on chitosan [□], chitosan-thioglycolic acid [◇], chitosan-thioglycolic acid with 426  $\mu\text{mol/g}$  disulfide bonds [▲] and 559  $\mu\text{mol/g}$  [●] disulfide bonds. Indicated values are means  $\pm$  SD ( $n \geq 3$ ). Differs from chitosan nanoparticles,  $p < 0.05$ .

# Content



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# Conclusion



- intravesical drug delivery system based on thiolated chitosan offers an adequate release profile besides its mucoadhesive properties
  - chitosan-TGA NP showed:
    1. greater stability
    2. superior mucoadhesion
    3. more sustained and controlled release
- Finally, chitosan-TGA intravesical drug delivery system might be a useful tool for a local drug application in the urinary bladder, which allows:
1. prolonged residence time at the target site
  2. enables sustained drug delivery of trimethoprim over a longer time span





**THANK YOU FOR  
ATTENTION!**