

## Aqueous Biopolymeric Coatings for Corrosion Protection of Galvanized Steel



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| Introduction  | Methodology   |
|---|---|
| Organic coatings are amongst the most popular   | 1. Solution (150 g/L) of Kraft Lignin (KL) in diethylene glycol monobutyl ether (DEGBE)       |
| surfaces [1]. However, the protection of metallic surfaces [1]. However, the production of these coatings is fundamentally dependent on petroleum | 2. Solvent exchange in water (1:10 Volumetric ratio of solution:water) $\longrightarrow$ CLPs |
| resources, resulting in materials with slow degradation<br>rates that accumulate as environmental pollutants [2].                                 | 3. TOCN suspension (1 or 2 g/L) + CLPs dispersion (1:1 Volumetric ratio)                      |
| Consequently, there is a demand for organic coatings  | = T + T + T + T + T + T + T + T + T + T   |

that are prepared from sustainable/renewable resources.

Lignin is the second most abundant biopolymer on earth and is the main constituent of the waste stream of wood-processing industries [3]. The combination of lignin and cellulose in composite films/coatings previously demonstrated promising barrier performances against water and oxygen permeation [4, 5].

Following a previous study on the performance of lignin as an anticorrosion coatings [6], this work investigated the corrosion protection capability of a water-borne composite coating based on colloidal lignin particles (CLPs) and TEMPO-Oxidized cellulose nanofibrils (TOCN) for galvanized steel [7].



4. Electrophoretic deposition (EPD, 0.5 or 3 V)

5. Drying and characterization of coatings (SEM, Cross-cut adhesion, EIS)



Figure 3. Optical microscopy images of coated steel after cross-cut adhesion measurements. (a) Coatings deposited at 0.5 V with 1 g/L TOCN concentration (0.1 T–0.5 V), and (b) at 3 V with 2 g/L TOCN concentration (0.2 T–3 V).

Table 1. Dried mass and thickness of coatings.

Figure 1. SEM micrograph of CLPs after drying on a silicon surface. Lignin particles demonstrated spherical morphologies and underwent coalescence during drying. The coalescence is resulted from the action of the utilized solvent (DEGBE).





Figure 4. EIS Bode plots of coated steel surfaces after 1 day (a) and 15 days (b) of immersion in 3.5 % NaCI.

Table 2. Charge transfer ( $R_{ct}$ ,  $k\Omega \cdot cm^2$ ) values obtained from fitting of EIS data.

| Sample   | 1 day immersion | 15 days immersion |  |
|--|-----------------|-------------------|--|
| HDG steel  | 0.9             | 0.2               |  |
| 0.1 T-0.5 V  | 27.0            | 6.0               |  |
| 0.2 T-3 V  | 34.0            | 13.7              |  |
| Conclusions  |                 |                   |  |
| <ul> <li>CLPs with a coalescing characteristic could be prepared.</li> <li>Electrophoretic deposition could be utilized for the preparation of aqueous coatings.</li> <li>Adhesion of TOCN-CLPs coatings on galvanized steel was affected by TOCN concentration and</li> </ul> |                 |                   |  |

Figure 2. (a) The schematic of electrophoretic deposition (EPD) of negatively charged biopolymers (CLPs shown as brown spheres and TOCN as gray lines), and (b) appearance of a biopolymeric coating immediately after the EPD process (2.5 cm × 4 cm dimension of steel substrate).

• Coatings provided a relatively long-term corrosion protection for the steel surface.

## References:

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deposition potential.

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